E 174 – DOE for Adipogenic Media Optimization

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Overview of Module 2

Two objectives

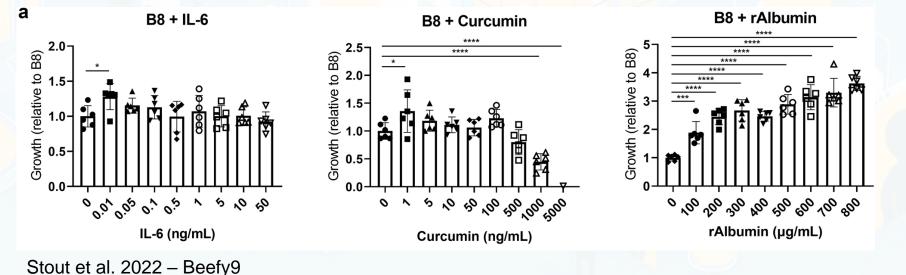
- To optimize the lipid accumulation media with design of experiments
- To grow a larger amount of pig adipocytes then aggregate them into tangibly large cultured fat tissues/constructs
- Ideally, we'd first optimize media then use that for the fat tissue construct
- But because of time constraints, these two objectives will be performed separately



What are strategies to optimize media?

One-factor-at-a-time (OFAAT)

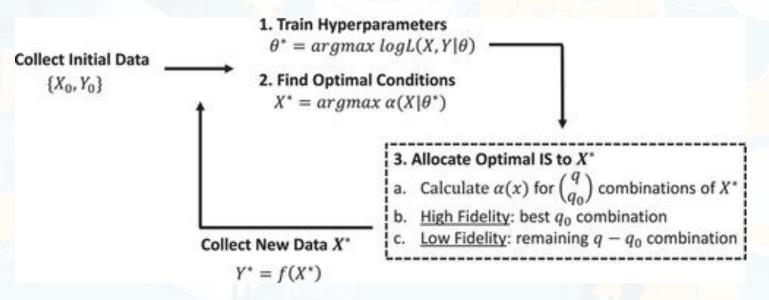
- Vary one factor (variable) while holding all others constant
- Easy design and interpretation
- Misses out on interaction between factors and is rather inefficient
 - → may miss the optimal set of conditions



What are strategies to optimize media?

Bayesian Optimization

- Global optimization of "black-box" function
- Complicated to understand

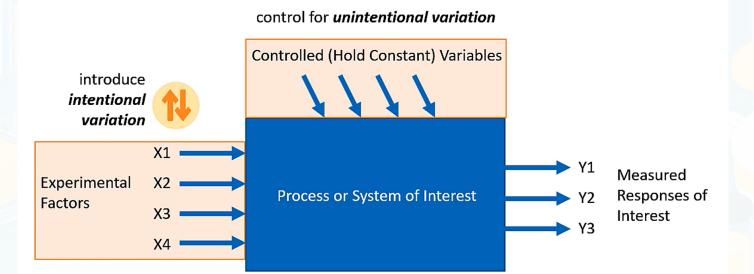


Cosenza et al. 2022. -- Multi-information source Bayesian optimization of culture media for cellular agriculture

What are strategies to optimize media?

Design of Experiment (DOE)

- Structured approach for data collection and making discoveries
- Flexible in that you can choose the complexity of the design and thus interactions



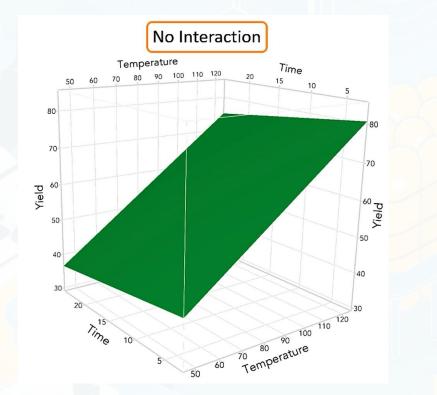
JMP statistical thinking for problem solving

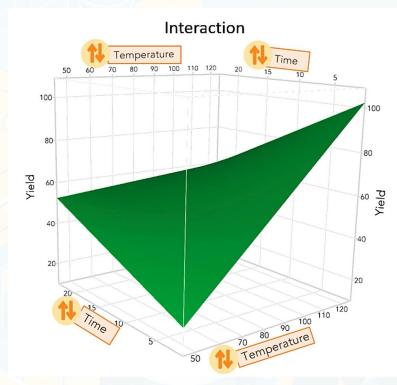
DOE: Interactions add curvature

Design space: Possible combinations of variables for experiments

Factors: Independent variables (experimental conditions)

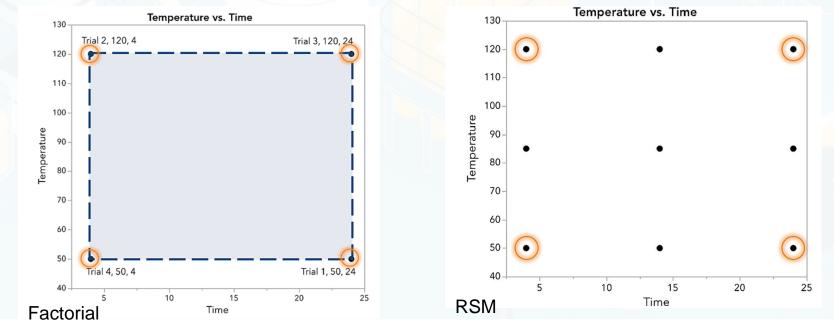
Response(s): dependent variables (what's measured)



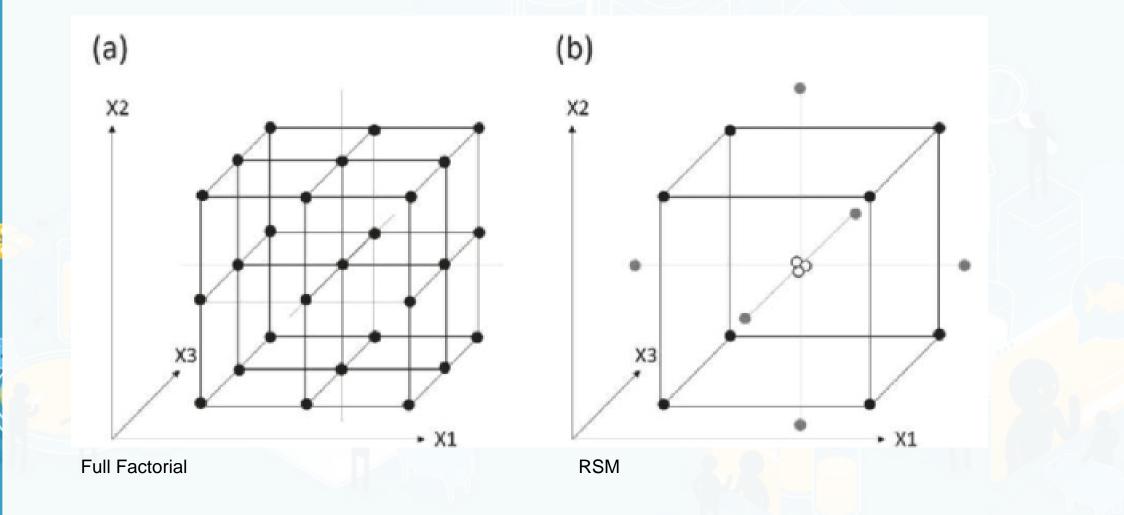


DOE Designs

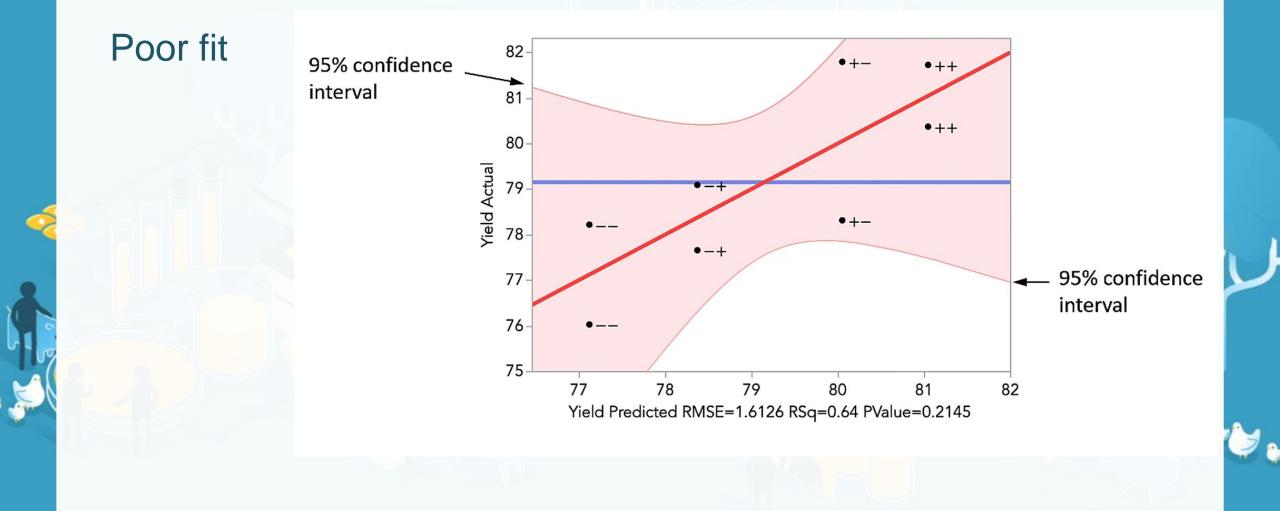
- General process
 - Screening design lots of factors, limited interactions
 - Full factorial design every possible interaction, complicated with higher number of factors
 - Response surface methodology (RSM) corners, "side" points, and center



Full factorial designs are more intense



DOE Analysis: Fit with model

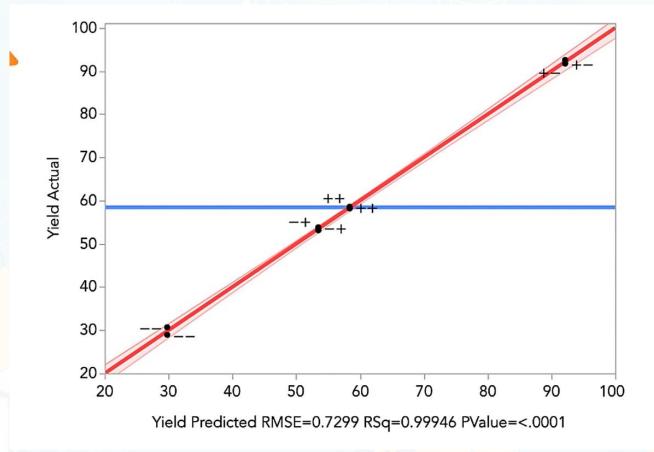


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DOE Analysis: Fit with model

We will be using standard least squares regression for our DOE analysis

Good fit





Class DOE

• As a team, we will contribute to a 4-factor DOE design

- \rightarrow 25 different media
- Every group will make 7 media (6 unique, one center point)
- We will be able to understand the interaction of these factors and model quadratic effects

Class DOE

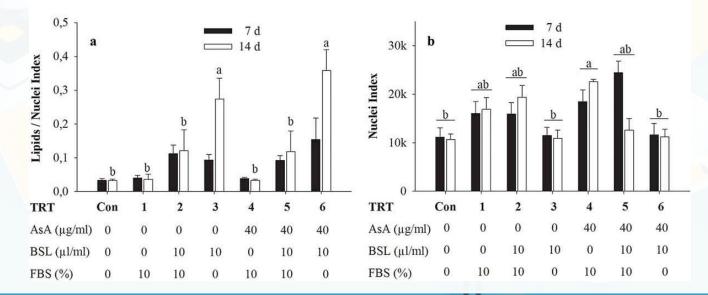
- We will optimize the best performing lipid accumulation regimes from Module 1 – DM2 from Jurek et al.
- As a reminder of the formulations
 - Induction Media (2 Days): DMEM, 10% FBS, biotin (10 μM), pantothenate (5.67 uM), insulin (3 μg/mL), dexamethasone (0.3 μM), IBMX (0.1 mM), rosiglitazone (10 μM)
 - Accumulation Media (Rest of the time): DMEM, 0% FBS, insulin (3 μg/mL), biotin (10 μM), 113 uM ascorbic acid, 500 ug/mL Intralipid



Class DOE

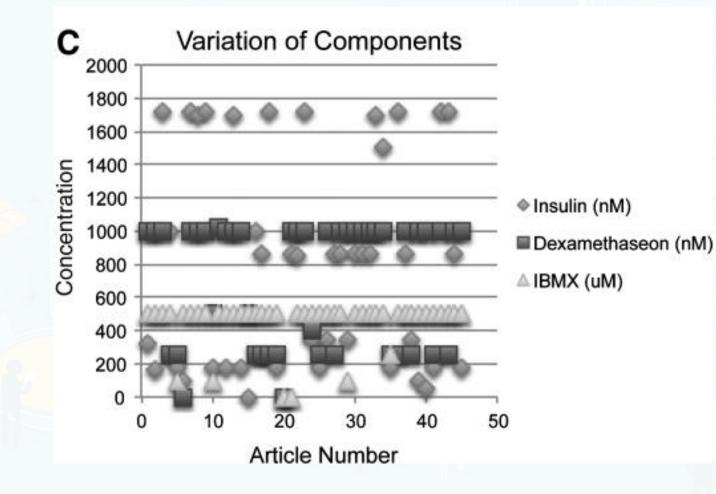
We will optimize the DM2 accumulation medium

- Jurek et al. tested ascorbic acid, bovine serum lipids, and FBS
- We will test additives of pantothenate and IBMX
- Also test varying concentrations of insulin and ascorbic acid



https://www.tandfonline.com/doi/full/10.1080/21623945.2020.1720480

Different Adipogenic Media Use Different Concentrations of Agonists

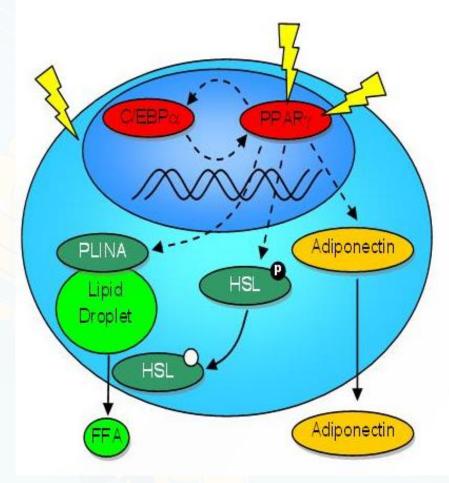


Which formulation is most optimal?

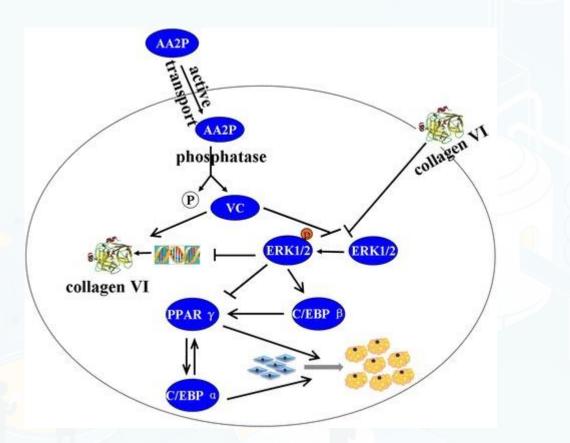


PPARγ is the master regulator of adipogenesis

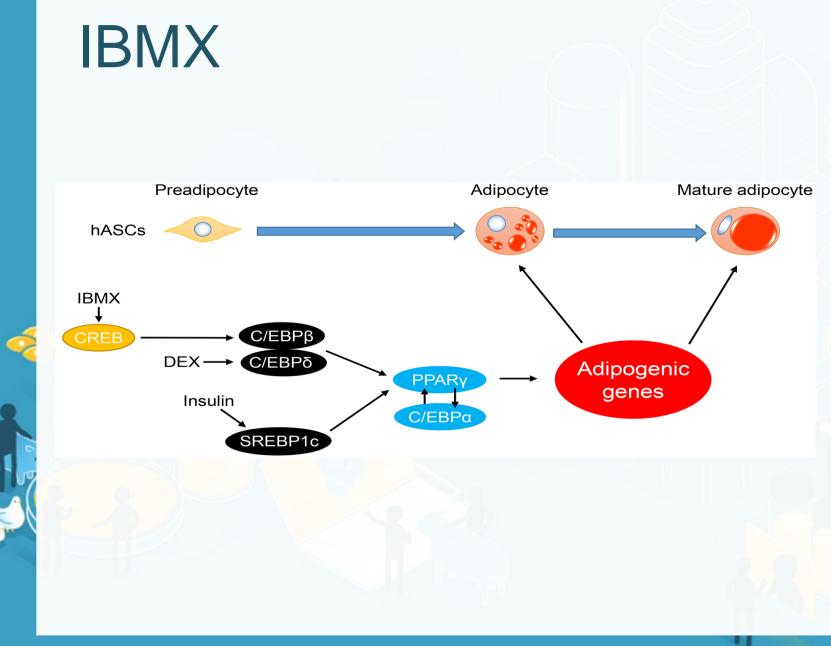
- PPARγ directly or indirectly up-regulates hundreds of adipocyte-specific proteins, which include
 - Adipogenesis regulators (C/EBPβ)
 - Lipolysis regulators (HSL)
 - Secretory proteins (adiponectin)



Ascorbic Acid



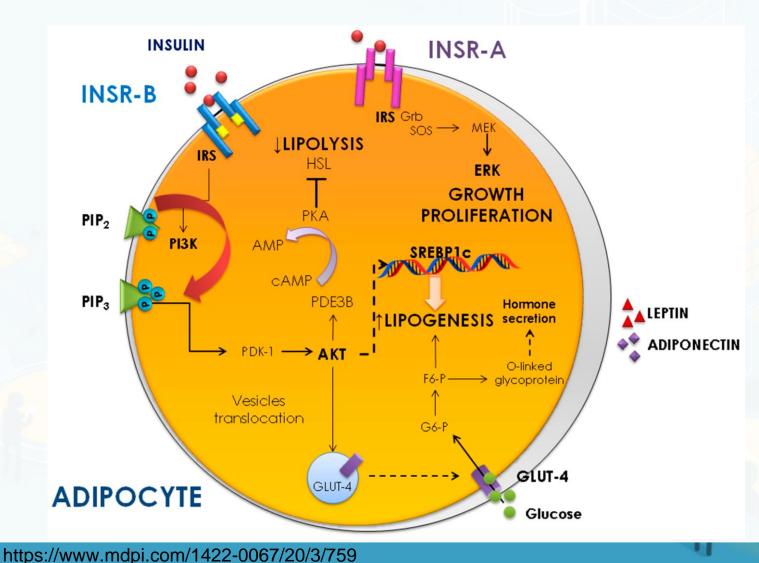
- Ascorbic acid treatment reduces ERK1/2 phosphorylation and increased collagen VI and adipogenic-specific factors
- ERK1/2 phosphorylation usually increase cell proliferation
 - ERK1/2 inhibits PPARy
- Collagen VI enriched in mature adipocytes



- Competitive, nonselective phosphodiesterase inhibitor, raising intracellular cAMP and protein kinase A (PKA)
- Stimulates CREB which in turn activates C/EBPβ
- C/EBPβ then activates PPARγ, the master regulator of adipogenesis



Insulin

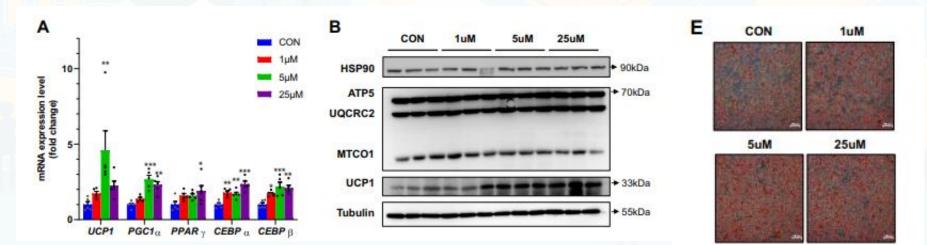


- Cross-talk with PPARγ
- Stimulates SREBP-1 and other transcription factors involved in adipogenic differentiation
- Limits lipolysis by inhibiting hormonesensitive lipase (HSL)

Pantothenate

Associated with brown adipose tissue activation

- Brown fat breaks down blood sugar (glucose) and fat molecules to create heat and help maintain body temperature
- White fat stores energy
- Pantothenate leads to dose dependent increase of UCP1, mitochondrial carrier protein found in brown fat



https://www.sochob.cl/web1/wp-content/uploads/2022/07/Pantothenate-protects-against-obesity-via-brown-adipose-tissue-activation.pdf

Response Surface Design

Made in JMP software

(Available for free through Tufts, but you don't need it for this class)

	Pattern	Insulin conc. (ug/mL)	Ascorbic acid (uM)	Pantothenate (uM)	IBMX (uM)	ORO Quantificatin (a.u.)	
Response Surface Design	1	0	0	0	0	•	
Responses	2+	0	0	0	200	•	
Add Response Remove Number of Responses	3+-	0	0	11.34	0	•	
Response Name Goal Lower Limit Upper	im 4++	0	0	11.34	200	•	
ORO Quantificatin (a.u.) Maximize .	5 -+	0	226	0			
	6 -+-+	0	226	0	200	•	
	7 -++-	0	226	11.34			
	8 -+++	0	226	11.34	200	•	
Factors	9 +	6	0	0			24 medi
	10 ++	6	0	0	200		for
Name Role Values Insulin conc. (uq/mL) Continuous 0 6	11 +-+-	6	0	11.34			
Ascorbic acid (uM) Continuous 0 226	12 +-++	6	0	11.34			
Pantothenate (uM) Continuous 0 11.34		6	226	0			and side
▲IBMX (uM) Continuous 0 200	14 ++-+	6	226	0	200		of
Factors	15 +++-	6	226	11.34			
ractors entral Composite Design	16 ++++	6	226	11.34			tessera
Display and Modify Design	17 a000 18 A000	0	113	5.67	100 100		(4D
Design Evaluation	19 0a00	3	0	5.67	100		version
Dutput Options	20 0A00	3	226	5.67	100		
Run Order: Keep the Same 🗸	20 0A00	3	113	0			cube)
Make JMP Table from design plus	21 00a0 22 00A0	3	113	11.34			
Number of Center Points: 4		3	113	5.67	0		-
Number of Replicates: 2	23 000a 24 000A	3	113	5.67	200		
Make Table	25 0000	3	113	5.67	100		- Fach
Back	26 0000	3	113	5.67	100		Each
Dack	27 0000	3	113	5.67	100		🔶 \succ group
	28 0000	3	113	5.67	100		center

point

replicate

Your media optimization

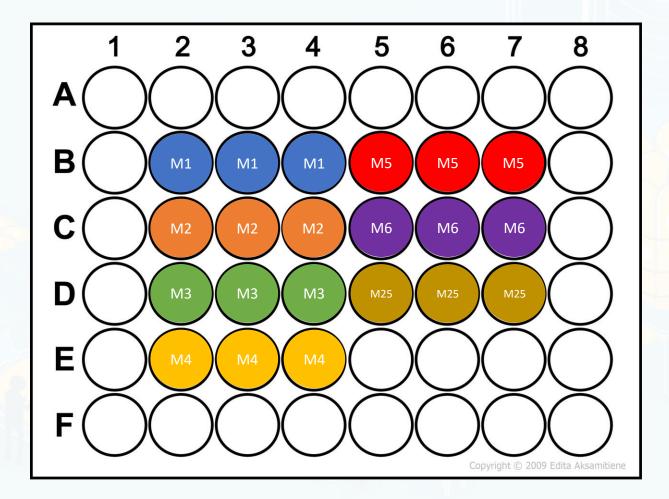
	Pattern	Insulin conc. (ug/mL)	Ascorbic acid (uM)	Pantothenate (uM)	IBMX (uM)	ORO Quantificatin (a.u.)	
1		0	0	0	0	•]
2	+	0	0	0	200	•	
3	+-	0	0	11.34	0	•	
4	++	0	0	11.34	200	•	
5	-+	0	226	0	0	•	
6	-+-+	0	226	0	200	•	
7	-++-	0	226	11.34	0	•	
8	-+++	0	226	11.34	200	•	
9	+	6	0	0	0	•	24 media for
10	++	6	0	0	200	• (
11	+-+-	6	0	11.34	0	•	corners and
12	+-++	6	0	11.34	200	•	sides of
13	++	6	226	0	0		tesseract (4D
14	++-+	6	226	0	200	•	
15	+++-	6	226	11.34	0	•	version of
16	++++	6	226	11.34	200	•	cube)
17	a000	0	113	5.67	100	•	,
18	A000	6	113	5.67	100	•	
19	0a00	3	0	5.67	100	•	
20	0A00	3	226	5.67	100	•	
21	00a0	3	113	0	100	•	
22	00A0	3	113	11.34	100	•	
23	000a	3	113	5.67	0	•	
24	000A	3	113	5.67	200	•	
25	0000	3	113	5.67	100	. ~	Each group's
26	0000	3	113	5.67	100	•	
27	0000	3	113	5.67	100	•	├─ center point
28	0000	3	113	5.67	100	•	replicate

- You will make 7 media
 - 6 unique ones (we will assign)
 - 1 center point (repeated by all groups)
- -1/a: lower bound of the variable
- 0: middle between lower/upper
- 1/A: upper bound of the variable



+ 2 additional replicates

Your well plate design



Different groups will be assigned different media

- Group 1: #1-6, 25
- Group 2: #7-12, 26
- Group 3: #13-18, 27
- Group 4: #19-24, 28



Module 2 Notes

THAWING: POOLING ALL GROUPS' CELLS TOGETHER
WE WILL USE PDFAT CELLS DURING THIS MODULE
DO NOT USE THE MEDIUM THAT CONTAINS PUROMYCIN

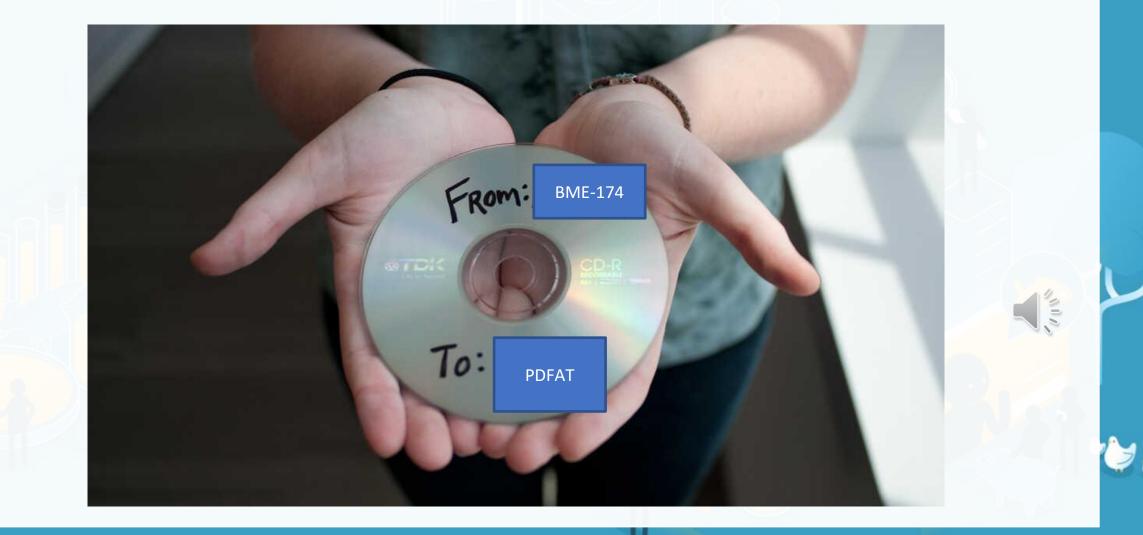


What adjectives describe Module 2 / Lipid Accumulation?

• Fatty



(Optional Assignment) Module 2 Mixtape



Smooth like ...



و 😂

Decadent / Rich



0

Affectionate



Euphoric / Glutinous

