#3 Biweekly Engineering Design Report

Project Title: Antibody Conjugated Silk Nanoparticles as a Targeted Treatment for

Glioblastoma Multiforme (GBM)

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Things highlighted in this color are new!

Project Description:

Glioblastoma Multiforme (GBM) is an aggressive tumor initiated by mutated astrocytes that can be found in the brain and spinal cord. As of now, the current treatment options for GBM are mainly surgery, radiation, and chemotherapy. These are all invasive or have severe side effects, so a targeted delivery system for chemotherapy using antibody-conjugated silk nanoparticles would be an important avenue to explore. The dual use of antibodies that target EGFRviii and IL-13Ra2 receptors are of interest. EGFRviii is a receptor expressed on the surface of around 30% of GBM cells, and not expressed in healthy brain tissue; IL-13Ra2 is expressed on 75% of GBM cells, but low level expression is found in the brain. The goals of this project are to determine the best receptors to target for GBM, determine the appropriate nanoparticle size for tumor uptake, induce successful antibody conjugation to the silk nanoparticle surface, and determine the proper antibody quantity required to have the nanoparticle be attracted to targeted receptors expressed by U87 cells.

Engineering Design Elements:

- What are the objectives of the project and the criteria for selecting them?
 - The objective of the project is to use antibody-conjugated silk nanoparticles as a potential method of targeted delivery for the treatment of Glioblastoma Multiforme (GBM). We aim to develop a 3D GBM model by seeding silk sponges with U87 cells that are transfected with a mutation of choice to carry the receptor we plan to target. We also plan to formulate silk nanoparticles of an appropriate size, loaded with doxorubicin, and conjugated with our antibodies of choice. We will add these nanoparticles to the 3D model and conduct imaging techniques such as live dead assays to quantify and analyze the efficacy of the drug delivery system. We also plan on validating antibody attachment to the particle surface using western blots, ELISAs, and fluorescent tagging. These objectives were chosen as they are crucial steps for developing and testing the nanoparticles to evaluate their potential and efficacy as a future treatment. As of now, the current treatment options for GBM are mainly surgery, radiation, and chemotherapy. These are all invasive or have severe side effects, so a targeted delivery system for chemotherapy would be an important avenue to explore for this unmet need, slowing down disease progression or relapse while decreasing major side effects.
- What system, component, or process is to be designed?
 - We plan on formulating a dual IL-13 and anti-EGFRviii conjugated nanoparticles to target GBM tumor cells. These antibodies bind to IL-13Ra2 and EGFRviii respectively, which are both expressed on the surface of many mutated GBM cells and have little to no expression in healthy brain tissue. Since different receptors

have been found to have varying levels of expression on healthy tissue and GBM cells, the dual targeting of two receptors is of interest. We determined the best antigen combination to target is IL-13Ra2 and EGFRviii. While IL-13Ra2 has one of the highest expression rates in patients, it is found in healthy brain tissues at low levels. EGFRviii, on the other hand, is only found on GBM cells, though it is only present in 20-30% of patients. The combined use of receptors will reduce off-target interactions and increase the GBM cells targeted in patients. The nanoparticles will be fabricated to be within a 100-120 nm size range, and they will be loaded at about 100-300 ng/ml concentration of doxorubicin¹. Furthermore, another component that must be designed is a 3D GBM model. U87 cells, which are human derived glioblastoma cells, will be transfected to express IL-13Ra2 and EGFRviii receptors. While many wildtype U87 cells already express these receptors, the population is so heterogenous that it will be most efficient for our project to ensure greater numbers of cells express our target receptors to test for efficacy. For our project, we will design a protocol to conjugate antibodies to the silk nanoparticle. It will likely be done using EDC/NHS, a crosslinking technique we have learned about through literature review. EDC, in conjunction with NHS, allows for a 2-step coupling of two proteins without affecting the carboxyls of the second protein. An alternative to EDC/NHS is coating the NPs with the antibodies, a process that would involve incubating the NPs in the antibodies diluted with PBS to induce tagging to the particle surface. Through literature review, past studies have used flow cytometry to calculate and validate the success of antibody-NP surface conjugation².

- What need does it fulfill (clinical, research, etc)?
 - As of right now, the current treatment options for GBM are mainly surgery, radiation, and chemotherapy. These are all invasive or have severe side effects, so a targeted delivery system for chemotherapy would be an important avenue and unmet need to explore slowing down disease progression/relapse while decreasing side effects.
- What scientific, math, and/or engineering methods will be applied?
 - Some of the scientific and engineering methods that need to be applied are silk processing, nanoparticle formation, antibody conjugation, doxorubicin loading of the nanoparticles, silk sponge formation for 3D brain model testing, and cell culturing. We will also likely employ western blots, ELISAs, fluorescent tagging, or mass spectrometry in order to detect the presence of desired proteins and antibodies.
- What realistic constraints (cost, safety, reliability, aesthetics, ethics, and social impact, etc) are to be considered?
 - One of the realistic constraints is understanding the right receptor to target; currently, our group is leaning towards a dual-targeting nanoparticle drug delivery technique, combining IL-13Ra2 with EGFRviii. We will only be relying on literature reviews for this decision, as purchasing three different antibodies and their transfection agents would be overly time consuming and expensive. Known limitations of targeted antibody therapies include off-target interactions. By researching and choosing antibodies with low-level expression outside of GBM tumors, we can decrease unnecessary exposure to chemotherapy. Finally, the U87

cells we are planning on using require transfection for the expression of biomarkers commonly found in patient tumors. None of us have ever performed viral transfection before, so this is a new area requiring training and certificates from the Biosafety Office. If this is needed for our project this year, we will need GFP/RFP expressing cell lines, for example. Furthermore, another constraint is time--there are few papers addressing dual-antibody conjugation, so the ability to successfully do so and develop techniques to evaluate our experiments may exceed our time limit of one school year.

- What alternative solutions or changes to the plan will be considered?
 - Our group decided that GBM was the best way to proceed onwards for now. Initially, we had only thought to conjugate just one antibody; our group's most recent alternative solution is to proceed with a dual targeting nanoparticle drug delivery approach and technique, combining IL-13Ra2 with EGFRviii. This will allow us to target a greater population of GBM cells while reducing off-target interactions. If we are unable to get lentivirus certification, instead of transfecting U87 cells to get higher expression levels of targeted receptors, we could use cell sorting to isolate cells with IL-13Ra2 and/or EGFRviii expression and culture them from there. We are also now in contact with Olivia's mentor from the City of Hope in potentially getting already-transfected U87 cells so we can bypass all transfection. If EDC/NHS is unsuccessful, we will look into simply coating the nanoparticles in the antibodies.
- What are the planned tests and what are the quantitative milestones that will demonstrate achievement of the objectives?
 - Some of the milestones and results that we have achieved so far include learning how to process silk and culture U87 cells in the Kaplan Lab (led by one of our group members Maddie) as well as learning how to make silk nanoparticles from Sunny (our lab mentor). A future quantitative milestone would include a protocol that has reproducible significant efficacy in producing either IL-13Ra2 and EGFRviii OR IL-13Ra2 and EphA2 antibody-conjugated silk nanoparticles.

Project Design Chart

Characteristic	Target Value	Why This Value	How We Will Test
Nanoparticle size	100-120 nm	Appropriate size for entering tumors via leaky vasculature and for tumor cell uptake	DLS/SEM imaging
Nanoparticle antibody expression	TBD → enough to have efficient uptake in GBM cells	Throughout various experiments, we will determine the target value for nanoparticle antibody expression based on which values optimize cellular uptake	FTIR Analysis, Fluorescence microscopy with secondary antibody

Silk concentration	6%	6% silk has been determined by past studies to result in 100-120 nm particles	Concentration calculations by weighing 1000ul of silk solution, leaving overnight in 60°C oven, and weighing remaining silk			
Uptake efficiency	TBD → enough to have efficient uptake in GBM cells	This value will be dependent on the various experiments we conduct to test nanoparticle antibody expression uptake efficiency (uptake is changed a lot by cell line & nanoparticle size ³)	FITC and lysosomal fluorescent microscopy or flow cytometry			
Cell receptor expression	Cells express one of each receptor	This is important to test the efficacy of dual antibody conjugation if both receptors are targeted	Flow cytometry and/or Western blot			

Specific Aim 1:

Develop dual antibody conjugated doxorubicin loaded silk nanoparticles Determine appropriate size, dose, and target receptors via lit review

Create product:

- Process silk and load with doxorubicin during formation of nanoparticles (100-120nm)
- Conjugate IL-13Ra2 and EGFRviii to loaded NP surfaces using EDC/NHS crosslinking

Characterize nanoparticles:

- Quantify ratio of antibodies on NPs via fluorescent imaging
- 2. Ensure successful crosslinking of antibodies

Specific Aim 2:

Create 3D GBM model using transfected U87 cells Transfect U87 cells to express the two receptors of interest:

- 1. Infect cells with lentivirus carrying genes for receptors
- Characterize cells to ensure receptor expression using western blot or ELISA

Seed transfected cells on silk sponges

Specific Aim 3:

Characterize therapy efficacy

Load nanoparticles onto the 3D GBM model:

- 1. Evaluate binding efficacy and nanoparticle uptake
- 2. Evaluate conditions of drug release
- Measure levels of apoptosis using live/dead assay

Compare efficacy of dual-antibody, antibody A, and antibody B nanoparticles

This flowchart describes the design of the proposed experiment. We will develop the dual antibody conjugated silk nanoparticles using established protocols used by the Kaplan Lab and EDC/NHS protocols published online, and we will characterize the nanoparticles produced. We will also transfect U87 cells to express our target receptors. These cells will be cultured, then later seeded onto



silk sponge scaffolds to create a 3D GBM model. Together, specific aims 1 and 2 will be used in conducting specific aim 3. The antibody tagged nanoparticles will be loaded onto the 3D GBM model and its performance will be evaluated. The efficacy of dual antibody and single antibody NPs will be compared to determine the best antibody coating for GBM targeting.

• Project Timeline:

https://docs.google.com/spreadsheets/d/1_7PQyerNKIqvtC1rLekw4sH9gVnIw_k4sOVCxkgSeg/edit#gid=0

• Specific Aims:

- 1. Develop dual-antibody conjugated silk nanoparticles
 - a. In order to test for the feasibility and efficacy of the project, the therapy must first be developed. Based on literature reviews and past studies, we aim to have a nanoparticle size of 100-120 nm and IL-13Ra2 and EGFRviii will be dually conjugated to particle surfaces. Established protocols are already in use at the Kaplan Lab for processing silk and measuring the size of the nanoparticles for characterization. Chosen antibodies will be conjugated onto the surface of the NPs using EDC/NHS protocols from past studies, and further testing will be conducted to determine the best way to conjugate two different types of antibodies at a constant, optimizable ratio. EDC/NHS is the conjugation method of choice as it is the most prevalent in previous literature, but we are planning on reaching out to experts in the lab who have performed antibody conjugation before to see what method they suggest before determining what protocol to use. We hope to fabricate nanoparticles with an antibody IL-13Ra2:EGFRviii ratio in three groups--1:1, 2:1, and 1:2. High resolution fluorescent imaging will likely be used to quantify antibody conjugation on individual NPs and quantified using ImageJ. This is necessary to ensure reproducible targeting effects and to better understand the formation of this therapy of interest. Successful completion of this aim will conclude in an established protocol for consistent dual-antibody conjugated nanoparticles.

2. Create a 3D GBM model using transfected U87 cells

We hypothesize that a 3D model of GBM will allow us to get the best in vitro observations of treatment efficacy and behavior. U87 cells are malignant glioblastoma cells; however, because they are a heterogenous population, it is best to transfect the cells to express the receptors of choice (biomarkers) so that they can be targeted by the antibodies. The U87 cells will be infected with a viral vector carrying the DNA sequence for either receptor and cultured. After cells are cultured and passaged, cells can be tested for receptor protein expression using western blots or flow cytometry. We hope to obtain a significant concentration of receptor protein when testing with western blots, and find that at least 60% of the U87s express one or more of the transfected receptors. Binding affinity of receptors to the purchased antibodies can be evaluated using flow cytometry, which will suggest the ideal receptor to antibody (therapy) ratio for best binding and interaction. Once they proliferate to a certain number, the transfected U87s will be seeded onto a silk sponge scaffold to form a 3D model and used for further testing. The expected outcome of this aim is a 3D GBM model with U87 cells that express target receptors that can bind to the purchased antiboies. Completion of

- this aim will produce a U87 culture transfected with receptors of interest to use in later experiments
- b. Instead of having to go through the process of virally transfecting U87s, there is the possibility of receiving cells from City of Hope Hospital in Los Angeles that express both biomarkers of interest. We could also use patient derived cells from a German hospital collaborator as well.
- 3. Characterize nanoparticle therapy efficacy
 - a. Characterization of nanoparticle efficacy is extremely important as it validates the feasibility of the product as a potential therapy for GBM. Nanoparticles of three groups would be loaded onto the 3D GBM models developed in Specific Aim 2: IL-13Ra2 + EGFRviii, IL-13Ra2, and EGFRviii. The efficacy of the three groups would be compared in terms of nanoparticle uptake. Nanoparticle uptake will be evaluated based on fluorescently imaging nanoparticles (using FITC) and lysosomes, then superimposing the images for overlap. The uptake of the three groups will be compared to the uptake of nanoparticles with no antibody coating (control), and their success will be measured with an uptake efficiency of 50% or above. These are all important markers of nanoparticle success for targeting and drug delivery. Successful completion of this aim will establish the efficacy of the dual-conjugated antibodies and confirm successful binding.
- What else is going on in the field that would compete with the project plans?
 - Something interesting going on in the field that could compete with the project plans is that some researchers were able to test silk fibroin nanoparticles coated with Tween-80 in GBM cell lines and found that they were able to release doxorubicin for up to 72 hours. Being able to cross the blood brain barrier is not necessarily something we must target in this capstone project, but it could be a future consideration to take into account if time permits. Our project also differs from this since ours would be more targeted due to antibody conjugation. https://www.sciencedirect.com/science/article/pii/S014181302034085X

Introduction and Background

Glioblastoma Multiforme (GBM) is the most common tumor in the central nervous system (CNS) and accounts for 65% of all CNS malignancies⁴. GBM is one of the most deadly forms of cancer, with a median survival rate of just 12.6 months after diagnosis⁵. Attributing to this severe prognosis is the tumor's location in the brain or spinal cord, severely limiting the success of traditional chemotherapies, radiation therapies, and surgical removal. Nanoparticles, however, are able to mitigate many of the obstacles that currently available therapies cannot overcome. Their advantages include biocompatibility, reduced toxicity, excellent stability, enhanced permeability and retention effect, and precise targeting⁶. The unique targeting ability of these nanoparticles can be enhanced with antibodies that bind to proteins on the surface of the selected cancer cells and deliver the drug of interest.

While nanoparticles can be composed of various materials, silk was selected as the appropriate material due to its biocompatibility, availability, and ease of size optimization and loading⁷. Further research will be done to determine the suitable size and loading dosage for the doxorubicin-loaded silk nanoparticles, for which we will follow an established protocol. Nanoparticles around 100 nm in the bloodstream are known to be too big to enter healthy tissue, but are able to enter tumors due to their leaky vasculature. Once they have entered the tumor and

bound to the cell receptors, they can be endocytosed to deliver the drug. Larger nanoparticles have been found to have longer rates of internalization; therefore, it may be advantageous to a NP large enough to only target cancerous tissue, yet small enough to be engulfed at an appropriate rate ⁸.

Epidermal growth factor receptors (EGFR) are transmembrane receptor tyrosine kinases (RTK) and are overexpressed in 50% of glioblastomas⁹. Epidermal growth factor variant three (EGFRviii) is a mutated wildtype EGFR expressed on the surface of GBM cells and commonly associated with GBM. This mutation has been found to lead to continued expression of tyrosine kinases, activate uncontrolled cell proliferation, growth, etc. EGFRviii is expressed in 25-33% of all GBM tumors in patients and it is not expressed in normal brain tissue ^{10, 11}. Some studies go so far to claim that EGFRviii has never been detected in healthy tissue¹¹. Its low expression in normal tissue makes it a suitable target for GBM therapies. Gliomas with EGFRviii have increased Ras activity, Akt/PI3k signaling, and increased expression of VEGF and IL-8¹². EGFRviii CAR T cells are in Phase I studies and have shown low off-target toxicity¹³.

Eph receptors are a class of tyrosine kinase receptors, and they are divided into A and B categories to indicate their extracellular domains. Subclass A indicates they are anchored to the membrane through glycosylphosphatidylinositol (GPI) linkage¹⁴. EphaA2 receptor was first discovered in 1990 and has since become a highly relevant receptor due to its abundance in several solid tumors^{14, 15}. EphA2 is over-expressed in ~60% of GBM tumors and is present in 98% of cells at moderate and or strong levels¹⁶. It is found to have a low-level expression in normal brain tissue. EphA2 is an attractive target since its associated with poor prognosis and survival due to its role in tumor cell proliferation, growth, and neovascularization ¹⁷. Functionally, EphA2 is a transcriptional target of the Ras-MAPK pathway¹⁸. Additionally, it has been found that in cancer cells, EphA2 receptors have ligand-independent kinase activity¹⁹.

Interleukin-13 receptor alpha2 (IL-13Rα2) was discovered as a glioma marker in 1995 by the Debinski laboratory²⁰, since then it has become one of the most studied tumor-specific antigens in glioblastoma research ²¹. IL-13Rα2 is a high-affinity membrane receptor of IL-13 and is expressed in many tumors²². It has been found to be overexpressed in up to 75% of glioma patients²³. Expression of IL-13Rα2 is high in the testis and placenta but has low expression in other organs²¹. A Phase III trial targeting IL-13Rα2 reported high levels of neurotoxicity due to off-target interactions with IL-13Rα1, a related receptor that is expressed in healthy brain tissue²⁴. While this trial revealed the dangers of working with IL-13Rα2, it suggests that there is promise if an antibody more specific to IL-Rα2 is found and used. Currently, CAR T-cell therapy targeting IL-13Rα2 is now in Phase I clinical trials ²⁵. Dual combinations of IL-13Rα2 and EphA2 have shown to be expressed in 90% of GBM patients} indicating promising data for better targeting specificity²⁶. EGFRviii and IL-13Rα2 targeted therapy has both been associated with recurrent antigen loss variants after initial treatment²⁷.

The combination of two receptor targets would allow for a greater number of GBM cell targets among its heterogeneous population, while also maintaining selectivity and reducing off-target interactions. Silk nanoparticles offer a unique opportunity to customize the drug, target, and dose of interest. In this project, doxorubicin loaded dual-antibody conjugated nanoparticles will allow for the targeted delivery of chemotherapy to GBM cells; compared to traditional therapies, the successful formulation of this therapy will result in more efficacious treatment for better patient outcomes.

Methods

Silk processing and nanoparticles formation are protocols from previous studies⁷.

Silk Processing⁷

Cut cocoons and remove inside layers, weigh out sodium carbonate to create a 0.02M sodium carbonate solution and add to distilled water. Add cocoon to boiling solution to degum silk fibers so that sericin is washed away and only fibrin protein remains. Wash degummed silk three times for 20 minutes each. Remove silk, pull by hand and air dry inside a fume hood. Add silk into 9.3 M LiBr solution to remove beta sheets and let sit for 4 hours at 60C. Inject silk LiBr solution into a dialysis cassette and remove air bubbles. Change dialysis water 3 times on the first day, twice on the second day, and once on the third day to wash out LiBr solution. Collect silk solution on day 3 and centrifuge solution twice, then store in the fridge for up to two weeks.

Silk Nanoparticles 7

5% silk solution will be added dropwise to acetone while maintaining a 75% acetone solution with doxorubicin in solution. The shear force of the droplets in the wall of the vortex will result in the formation of nanoparticles, and doxorubicin will be encapsulated within in the process. Precipitant will then be centrifuged at 48,000 g for 2 hours. Supernatant is aspirated and pellet is resuspended in 20mL of distilled water. Vortex and apply two sonication cycles at 30% amplitude for 30 seconds. Repeat centrifusion and resuspension 2> times. Resuspend pellet in distilled water and store for later use. The values mentioned in the above protocol can be optimized to produce the desired nanoparticle size as determined from literature reviews.

U87 Culturing

After taking 1mL U87 cell line out of the liquid nitrogen freezer and thawing in metal beads around 34C, 100 µL of that cell suspension can be added to 9.9mL of cell culture media in a T75 to grow for 2+ days until 80-90% confluency is reached, which is about 2-3 million cells in a T75. U87 cell culture media contains MEM, FBS, penicillin, and streptomycin. U87 cells are non-adherent and naturally like to grow in aggregates, making spheroids. To prevent this, cells are cultured on plates that have been coated with Poly-L-ornithine (PLO), which promotes cell adherence and long term culture of neuronal cells. After a few days, the cells will need to be processed, which involves suctioning out the old culture media, adding 2mL of trypsin (a digestive enzyme) to the T75, and incubating for 5-10 minutes at 30 C until the cells have lifted from the PLO plate. After incubation, the flask is visualized under the microscope to determine if the cells have lifted. Once this has occurred, 8mL of cell culture media is added to the cell suspension, and the 10mL solution is transferred to a 15mL tube where it is centrifuged at a speed of 400G for 5 minutes. After centrifugation, the cells are left as a pellet at the bottom of the supernatant, which is removed to leave the pellet. The pellet is resuspended in 1mL of culture media using a P1000 pipette. A T75 flask with PLO seeded is taken from the incubator to plate the cells. To plate at a lower density, 100 µL of cell solution is added to 9.9mL of media for a 1:10 ratio, while a 1:5 ratio would be 200 µL of cell solution with 9.8mL of media. The new T75 is placed back into the incubator until processing.

Results

We were able to successfully obtain 13 mL of silk solution, which we will be evaluating for concentration and using to formulate nanoparticles of 100-120 nm in diameter. The group also

knows how to culture U87s, so transfected versions of U87 cells expressing biomarkers IL-13Ra2 and EGFRviii will be used to demonstrate cellular uptake of the silk NPs.

Like many tumors, gliomas contain Next, antibodies for IL-13Ra2 and EGFRviii Glioblastoma Multiforme (GBM) is an (2)biomarkers of certain receptor and will be conjugated to the nanoparticle surface using the EDC/NHS protocol; aggressive tumor initiated by mutated protein mutations astrocytes that can be found in the brain and microscopy will be used to determine spinal cord These mutations can be used as homogeny between the conjugated NPs targets for drug delivery systems. like antibody conjugated nanoparticles First, silk nanoparticles will be constructed dropwise in an acetone solution (3) Finally, fluorescence microscopy will be used to Then, using either virally transfected U87s quanitify how successfully the antibody conjugated NPs were uptaken by GBM cells or patient derived cells expressing both antigens, antibody conjugated NPs will be added to the culture to stimulate cell

Dual Antibody-Conjugated Nanoparticles for Targeted GBM Treatment

We were also able to create a unifying figure for our entire project, which is a simple figure that provides a visual of our project: the problem of interest, objectives, and end goals.

Discussion

We processed silk two times this semester. The first time, the pH of the water was around 5, which affected the silk and resulted in unexpected visual cues after dissolving in LiBr. This prompted us to discard that batch and process silk a second time, where the dissolving and dialysis of the silk was much more routine. As stated in the results, we were able to obtain a 13 mL silk solution that is stored in the fridge and will soon be used for nanoparticle fabrication.

Participation: List individual contributions of each group member to the project

- Maddie Yost: GBM lit review research, antibody (EGFRviii) lit review research, lead silk processing and cell culture training for group, added to/edited Biweekly report and Midterm Technical Report/Presentation, Zoom meeting with Dr. Saul Priceman (PhD from City of Hope) who is an expert in the field for this type of research
- Olivia Zeiden: GBM lit review research, Breast cancer lit review (ultimately ruled out), met with maddie to learn silk processing, met with Sunny for silk nano particle training, added and edited Biweekly report, updated project timeline with relevant dates and aims, EphA2 Antibody lit review, added to the midterm report and presentation, added to midterm report #3, Zoom meeting with Dr. Saul Priceman (PhD from City of Hope) who is an expert in the field for this type of research.

- Sabrina Zhang: GBM lit review research, hepatocellular carcinoma (ruled out target) lit review, met with Maddie for silk processing and U87 cell culture training, met with Sunny for silk nanoparticle training, edited project schedule, wrote brief blurb for Sunny on the need for our proposed GBM treatment, added to/edited Biweekly report #1, lit review for IL-13Ra2 as potential target, added to Biweekly report #2 and Midterm Mid Semester Technical Report, Zoom meeting with Dr. Saul Priceman (PhD from City of Hope) who is an expert in the field for this type of research. Edited Biweekly report #3.
- Elysia Chang: GBM lit review research, met with Maddie and the group to learn silk processing and cell culture, met with Sunny to conduct silk nanoparticle training, added to/edited Biweekly report, created the project timeline, HCC initial research (ruled out target), EGFRv3 research to see if it is a good target, created Midterm Presentation file (because unable to present in-person/on Zoom due to the Society of Women Engineering 2022 Conference in Texas), created/added to Midterm Mid Semester Technical Report, Zoom meeting with Dr. Saul Priceman (PhD from City of Hope) who is an expert in the field for this type of research

Scoring Metrics

Project Description: 2 points

Engineering Design Elements: 8 points (1 point per question)

Advice - Structure the design reports so they evolve into your mid semester and final technical reports, as a living document to make your writing and reporting easier

Appendix 1 Project Schedule

Aims	Sub-Aims	Completion	September	October	November	December	January	February	March	April	May
Define Project		100									
1st Biweekly Report		100									
Antibody Lit Review	Choose Antibody for Targeting Research	100									
Silk Processing		100									
Silk Nanoparticles		100									
2nd Biweekly Report		100									
Cell Culture Training		50									
Culturing U87s		25									
Technical Proposal Report Draft		100									
Project Presentations		100									
Preparing Nanoparticles		50									
Start Website	Add in Home Page, People Sections, References, and Project Update	100									

3rd Biweekly Report		0					
Risk Assesment Analysis		0					
Update Website	Update Project Section and References	0					
Order Materials Necessary for Antibody Conjugation		0					
4th Biweekly Report		0					
Update Website	Update Project Section and References	0					
5th Biweekly Report		0					
Project Presentations		0					
Finalized Web Site		0					
Technical Report		0					

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